

# Molecular and epidemiological investigation of a colistin-resistant OXA-23-/NDM-1-producing Acinetobacter baumannii outbreak in the Southwest Indian Ocean Area

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#### Molecular and epidemiological investigation of a colistin-resistant OXA-

#### 23/NDM-1-producing Acinetobacter baumannii outbreak strain from-in

#### Southwest Indian Ocean

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#### Abstract

Dual resistance to colistin and carbapenems is a milestone reached by certain extensively-drug resistant (XDR) Gram-negative bacteria. The present study describes the first outbreak of XDR colistin- and carbapenem-resistant OXA-23/NDM-1-producing *Acinetobacter baumannii* (CCRAB) in European overseas territory of Reunion Island (France, Indian Ocean). Between April 2019 and June 2020, 13 patients admitted to the University Hospital of Reunion Island were involved in the outbreak, of whom 8 were infected and 6 died. The first case was traced to a medical evacuation from Mayotte Island (Comoros archipelago). An epidemiological link could be established for 11 of the patients. All of the collected CCRAB isolates showed the same resistance profile and co-produced intrinsic β-lactamases OXA-69 and ADC-191, together with acquired carbapenem-hydrolysing β-lactamases OXA-23 and NDM-1. A mutation likely involved in colistin resistance was detected in two-component system PmrAB (D82N in PmrA). All of the isolates were found to belong to ST<sup>Pas</sup>1/ST<sup>Ox</sup>231 clonal complex, and phylogenetically indistinguishable. Their further characterisation by whole genome sequence analyses (wgMLST, SNPs) provided hints about the transmission pathways. This study pleads for strict application of control and prevention measures in institutions where the risk of imported XDR-bacteria is high.

- Keywords: Acinetobacter baumannii; extensively drug-resistant; outbreak; colistin and carbapenem-
- 49 resistance; Reunion Island; Indian Ocean.

#### 1. Introduction

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Antibiotic resistance is a global threat to public health, with considerable clinical and economic impacts [1]. Carbapenem-resistant Gram negatives are the most challenging pathogens worldwide and among them, multidrug-resistant strains of Acinetobacter baumannii are increasingly reported in Europe [1,2]. While invasive carbapenem resistant A. baumannii strains resistant to carbapenems (CRAB) remain relatively infrequent in mainland France (9% in year 2018), they are more prevalent in neighbouring countries (average of 32.6%) [2]. Because of their spreading capacity within and between medical institutions, CRAB isolates represent a serious threat to fragile patients, notably those hospitalised in intensive care units (ICU). In Acinetobacter species, carbapenem resistance mostly results from horizontal transfers of genes encoding class D (OXA-23, -40, -58) or class B βlactamases such as NDM-1 [3]. Co-occurrence of several carbapenem-hydrolysing enzymes such as OXA-23 and NDM-1 has even been detected in strains from India and China back in 2010, and later on in European countries [4-7]. Because polymyxins often remain the ultimate therapeutic option against CRAB-associated infections, resistance to these agents has raised in institutions or geographic areas where the prevalence of CRAB is elevated [8,9]. As for several other Gram-negative pathogens, polymyxin resistance in Acinetobacter sp. A. baumannii is associated with mutations that activate a two-component system (PmrAB), with subsequent addition of phospho-ethanolamine to the lipopolysaccharide and a final decrease in net negative charge of the bacterial surface [10]. A first outbreak of carbapenem- and colistin-resistant A. baumannii was described in Spain in 2009 [11]. Other episodes were reported in Italy in 2014 and in Greece in 2015, but that they did not imply strains coproducing NDM-1 and OXA-23 [12,13]. Reunion Island is a French overseas territory of 850,000 inhabitants, located in the Southwest Indian Ocean Area (SIOA), close to Madagascar and to Mayotte Island (Comoros archipelago, about 250,000 inhabitants). It offers the same healthcare facilities as in mainland France, with the University Hospital of Reunion Island (URHI) being considered as the reference hospital in SIOA. Due to this sanitary situation and the large population flows between Reunion Island and both the Indian subcontinent and Southern Africa, Reunion Island is exposed to multiple and repeated imports of multidrug-resistant bacteria including carbapenemase producers. As an example, its epidemiology of Carbapenemase-Producing Enterobacteriaceae Enterobacterales-(CPE) is radically different from that of occidental Europe (dominated by OXA-48-like enzymes) since bland-positive isolates account for 79.4% of CPE [14]. Concerning Acinetobacter species, the first NDM-1<sub>7</sub> and OXA-23 coproducing isolates from SIOA, were detected in April and December 2015 in Mayotte Island and in Reunion Island, respectively (unpublished data from the French National Reference Centre for Antibiotic Resistance, FNRC-AR). According to the Federation for Nosocomial Infection Control of Reunion Island (FNICRI), the rates of CRAB in Reunion Island were 11.4% and 12.7% in 2017 and 2018, respectively [15].

Here, we describe the main findings related to an outbreak that occurred between April 2019 and August 2020, and that involved for the first time an OXA-23/NDM-1-positive, colistin-resistant *A. baumannii* (CCRAB) clone.

#### 2. Materials and methods

- 2.1. Study setting and data source
- 91 2.1.1. Study setting

This study was approved by the Ethics Committee of the French Society of Infectious Diseases, and was declared to the Commission Nationale de l'Informatique et des Libertés (French Data Protection Agency or CNIL, MR004). The need for informed consent was waived, as the study was non-interventional and followed our usual protocol. However, all patients or their legally authorised representatives were verbally informed about the process of data collection, and could refuse to

participate in the study. This study complies with the Strengthening the Reporting of Observational studies in Epidemiology recommendations statement [16].

#### 2.1.2. Data source

The UHRI is an 1,800-bed facility that generates 175,000 hospital stays and 315,000 outpatient consultations per year. It consists of two sites located in the Northern and in Southern parts of the island. The outbreak occurred in the Northern site of UHRI, in the intensive care and nephrology units. The ICU consists of 3 subunits, A, B and C, each containing 10 beds. In agreement with the institution hygiene policy, patients admitted to ICU are systematically screened for carriage of multidrug resistant microorganisms, and weekly thereafter.

During the outbreak investigation, the following data were collected for each CCRAB-positive case: age, gender and geographical origin of patients, hospitalisation ward and dates, room numbers, sampling dates (1<sup>st</sup> positive sample), specimen sources, infection or colonisation types and death (where applicable).

#### 2.2. Outbreak detection and investigations

#### 2.2.1. Outbreak detection

The outbreak was suspected on mid-July 2019 when 3 XDR *A. baumannii* strains were successively recovered over 5 weeks from 3 patients hospitalised in the same ICU of UHRI. The 3 isolates displayed identical antibiotic susceptibility profiles, and were referred to the FNRC-AR for confirmation of colistin resistance and genotypic comparison. A multidisciplinary team composed of epidemiologists, public health officials, ICU doctors and microbiologists was set up to control the outbreak, and to conduct epidemiological and environmental investigations.

#### 2.2.2. Outbreak investigations

The outbreak investigation comprised: (i) immediate risk assessment of procedures involving the use of equipment shared between the different ICU rooms, and determination of all contact patients; (ii) repeated rectal screening of contact patients (3 screenings one week apart); (iii) environmental sampling of the rooms of CCRAB positive patients, and of shared equipment; (iv) multidisciplinary meetings driven by the infection control team, with ICU medical doctors, hygienists and microbiologists to decide on the follow-up of the outbreak; (v) interviews of ICU teams to identify risk behaviours, and re-sensitisation of staff about contact precautionary measures to prevent cross transmissions. From March 2020, contact patients were additionally screened for pharyngeal and cutaneous (armpit) CCRAB carriage.

#### 2.3. Case definition and patient outcome

We defined as positive case, every (non-redundant) patient from whom at least one clinical sample cultured a colistin-resistant (minimum inhibitory concentration, MIC > 2 mg/L), carbapenem-resistant (MIC meropenem > 8 mg/L and/or imipenem MIC > 4 mg/L) *Acinetobacter baumannii* (CCRAB) strain, according to EUCAST guidelines [17]. Patients sharing the same paramedical team as those of a CCRAB carrier were considered as potential contact patients [18]. An epidemiological link was considered probable between CCRAB positive patients if those had stayed in a same ward, under spatiotemporal conditions compatible with a CCRAB transmission *via* the nursing staff, shared equipment, or room environment.

The clinical outcome of patients was followed for a period of 3 months after their discharge from the medical wards, where the CCRAB had been detected. The notion of attributable death was determined on a consensual basis by the multidisciplinary meeting group.

#### 2.4. Sample collection and laboratory investigations

#### 2.4.1. Bacterial isolates and antimicrobial susceptibility testing

Acinetobacter baumannii strains were identified by MALDI-TOF mass spectrometry (Microflex, Bruker Daltonics, Marne-la-Vallée, France) according to the manufacturer's recommendations, and further characterised by whole genome sequence (WGS) analysis. MICs of 19 relevant antimicrobials were determined by using gradient strips (Etest, bioMérieux, Marcy l'Étoile, France) or by the broth microdilution method (ThermoFisher Scientific, Courtaboeuf, France), notably for colistin. All MICs were interpreted according to EUCAST 2019 breakpoints [17].

#### 2.4.2. Whole genome sequencing and bioinformatic analyses

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Genomic DNA was extracted with the PureLink<sup>™</sup> Genomic DNA mini kit (ThermoFischer Scientific). Whole genome sequencing (WGS) was performed on the P2M platform (Institut Pasteur, Paris, France). In brief, Nextera XT DNA Library Preparation kit (Illumina) was used for library construction. The pooled libraries were sequenced on an Illumina NextSeq 500 platform using paired-end 150-bp runs. The fastq files were generated and de-multiplexed with bcl2fastq Conversion Software (v2.20 Illumina). The final average sequencing depth was around 80 X. The reads were assembled de novo by using SKESA (v2.4.0), and the contigs were annotated with Prokka (v1.14.5). Whole genome Multi Locus Sequence Typing (wgMLST) analysis was performed from annotated contigs by using PPanGGOLiN (v1.1.85) that defined 3,523 « persistent » genes, and then filtered by Gubbins (v2.4.1) for single nucleotide polymorphism (SNP) identification and filtering [19]. SNP analysis of the 14 CCRAB A. baumannii isolates was conducted by using the closest reference genome available (A. baumannii AR\_0083, accession number: CP027528) and Snippy (v4.4.5), with a minimum sequencing depth at SNP position equal to 10, and a minimum distance between SNPs equal to 11 bp. Reads mapping was carried out by using CLC Genomics Workbench (v10.1.1) (Qiagen, Courtaboeuf, France). The antimicrobial resistance genes were identified by uploading the sequence reads to the CARD website (https://card.mcmaster.ca). The sequence type (ST) of isolates was determined according to both Oxford and Pasteur MLST schemes (https://pubmlst.org/abaumannii/), thanks to using the online MLST 1.8 tool (https://cge.cbs.dtu.dk/services/MLST/). This Whole Genome Shotgun project PRJNA658235 was deposited at GenBank under the accession references JACORF000000000 to JACORS000000000.

#### 3. Results

#### 3.1. Outbreak description and demographic data

A total of 13 CCRAB positive patients were identified at the Northern site of UHRI between 29<sup>th</sup> April 2019 and 1<sup>st</sup> June 2020. An epidemiological link was established for 11 ICU cases. Two additional CCRAB positive patients (Case 1 admitted to the infectious diseases unit and Case 5 admitted to the nephrology ward) were considered as part of this outbreak despite the absence of obvious epidemiological link with the other 11 cases. CCRAB isolates were collected between June and August 2019 (phase II), November and December 2019 (phase II), February and March 2020 (phase III), and May and August 2020 (phase IV; Figures 1.a and 1.b). Nine of the 13 CCRAB positive patients (69.2%) were male, with a median age of 58 years (IQR: 45.5-66.5, Table 1). The mean length of ICU stay was 30.9 days (range: 3-97 days). Five patients were colonised and 8 patients infected, including 4 cases of bacteraemia, 3 cases of pneumonia (of which 2 became bacteraemia), and 1 case of osteoarticular hip infection. Of the 13 CCRAB cases, 6 died (46.2%) including 4 fatal outcomes considered as directly attributable to CCRAB infection (30.8%). All these 4 attributable deaths were caused by a refractory septic shock linked to ventilator-associated pneumonia (Cases 4, 9 and 13), and to bacteraemia (Case 11), respectively. Since the end of August 2020, no additional CCRAB strains were isolated in UHRI.

#### 3.2. Outbreak investigation and control measures

#### 3.2.1. Outbreak investigation

Retrospective epidemiological investigations showed that Case 1 had been hospitalised for 4.5 months at the Hospital Centre of Mayotte for a complicated infection of left hip, prior to his transfer

to UHRI. In Mayotte, he received multiple rounds of antibiotic treatments, but no colistin. Case 12, a patient living in Madagascar, was also medically evacuated to UHRI because of a septic shock subsequent to necrotizing fasciitis of his right leg. Before his transfer to Reunion Island, he had been hospitalised for 3 days in a Malagasy hospital, where he received several courses of antibiotics, but no colistin. No travel history was recorded for the other 11 patients.

Our retrospective analysis also revealed that Case 6 was hospitalised in the same ICU room as Case 3 (Figure 1.a). Case 7 was hospitalised in the same room as Case 4, 4 days after his death, while Case 3 and Case 4 shared the same consulting room (4D) six days apart (Figure 1.a). All patients rectal screenings were negative for *A. baumannii* at the time of ICU admission. At day-21 post-admission, Case 3 was detected positive for three carbapenemase-producing enterobacteria (each producing the OXA-181 enzyme) in addition to the CCRAB strain. Since March 2020, all patients with a positive CCRAB rectal screening were found colonised both in the pharynx and the armpit.

A total of 113 environmental samples (63 in ICU and 50 in nephrology unit) were collected between July 2019 and September 2020, from patients rooms (siphons, toilets, environment) and from the medical equipment shared between rooms (ultrasound scanner, electrocardiograph, tensiometer, patient lift, ...). All samples turned out to be negative by culture except one recovered in August 2019 from a chair located in the Case 7 room (Figure 1.a).

#### 3.2.2. Control measures

During the 3<sup>rd</sup> phase of the outbreak (March 2020 in ICU), three major measures were implemented to control the outbreak. Firstly, all ICU patients were screened simultaneously at three body sites (rectum, pharynx and armpit), and positive patients were segmented in subunit A, while new admissions were oriented to the two other subunits, B and C. A complete sequential decontamination (five rooms by five rooms) of subunit A was then implemented. Thirdly, a hygienist nurse of the infection control team was specially assigned to the ICU healthcare team to observe, to

advise and re-train the staff in preventive measures against cross-transmissions. The main difficulty encountered was to apply these decisions in emergency because the first case of SARS-CoV-2 infection had been detected two days earlier on Reunion Island, and all the ICUs had to organise dedicated subunits for SARS-CoV-2 positive patients.

During the 4<sup>th</sup> phase of the outbreak (June 2020 in ICU) with the resurgence of Case 13, a collegial discussion was held on whether or not to close subunit C. However, this decision was not taken because of the sanitary context due to SARS-CoV-2 pandemic, and the limited number of ICU beds available in Reunion Island and in SIOA.

#### 3.3. Laboratory investigations

#### 3.3.1. Clinical microbiological data

A total of 14 isolates of *A. baumannii* were collected, 13 from Cases 1 to 13, and 1 from Case 7 room (named E14). All exhibited the same high resistance to antimicrobials including imipenem, meropenem, cefiderocol, amikacin, ciprofloxacin and colistin, while remaining susceptible to tobramycin (MIC range: 0.5-1 mg/L). Tigecycline and rifampicin MICs ranged from 0.25 to 1 mg/L and from 3 to 4 mg/L, respectively (Supplementary table 1).

#### 3.3.2. Molecular and sequencing data

#### 3.3.2.1. Antibiotic resistance genes characterisation

WGS experiments analysis revealed that all the isolates possessed the intrinsic genes  $bla_{OXA-69}$  and  $bla_{ADC-191}$ , and shared the same resistome. Accounting for their pan resistance to beta-lactams, all the bacteria harboured the carbapenemase-encoding genes  $bla_{NDM-1}$  and  $bla_{OXA-23}$ . Gene  $bla_{NDM-1}$  was localised within an intact Tn125 transposon, whereas two copies of  $bla_{OXA-23}$ -carrying Tn2006 were detected in the bacterial genomes. Were also identified Resistance determinants to aminoglycosides (genes aadA1, aacC1, aphA6, aphA7, strA, strB), sulphonamides (sul1, sul2), trimethoprim (dfrA1),

tetracyclines (tetB, tetR), disinfectants ( $qacE\Delta 1$ ) and heavy metals (merRTPCADE operon) were also identified.

To get an insight into the mechanism of polymyxin resistance in the CCRAB strains, sequences of several genes known to be involved in colistin resistance were aligned with those of reference or clinical susceptible strains. No mutation were identified in the following genes: *lpxA*, *lpxC*, *lpxD*, *lpsB*, *lolA*, *lolB*, *pldA*, *ttg2E* (data not shown). *Mcr*-like genes were not detected in CCRAB isolates either. However, as compared with the sequence of *pmrCAB* operon from closely-related susceptible *A*. *baumannii* strains (same ward, same resistome, only 13 SNPs of difference with resistant strains), all CCRAB strains appeared to contain a D82N substitution in response regulator PmrA, whose relevance is currently studied.

#### 3.3.2.2. Genotyping data

All the CCRAB strains belonged to ST<sup>Pas</sup>1/ST<sup>Ox</sup>231, and are therefore affiliated to International Clone 1 (also referred to as Global Clone 1 or GC1). WgMLST method was used to better discriminate the isolates. Compared with other ST<sup>Pas</sup>1 *A. baumannii*, the 14 CCRAB isolates clustered together, differing from each other by 0 to 5 SNPs (Figure 2.a and supplementary table 2). More in-depth analysis of SNP calling was undertaken in order to provide some hints about the transmission events that occurred during the outbreak. This led to the identification of several subclusters (Figure 2.b). The first one involved two patients (Cases 6 and 12) with almost identical CCRAB isolates (differing by only 1 SNP) dating from July 2019 and March 2020. The fact that no temporal connection between the two cases could be found strongly suggests a role for the ICU environment in contamination of the second patient. Two other distinct subclusters were identified, each involving strictly identical isolates (Cases 3-4 and Cases 9-10-11, respectively), supporting a cross transmission between the corresponding patients. Finally, as expected, the environmental isolate recovered in Case 7 room was very close to that identified in Case 7 (Figure 2.b).

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#### 4. Discussion

XDR-Acinetobacter baumannii (XDR-AB) represent a growing concern in hospitals, particularly in ICUs, because of the ability of these bacteria to become resistant to any available antibiotics, and to persist in the environment [20–22].

Case 3 can be viewed as the index patient of this ICU outbreak, that initially started in subunit C and rapidly spread to subunit A (Figure 1.a). Concerning the acquisition of CCRAB by Case 3, two hypotheses can be considered, (i) a transmission from Case 1 to Case 3 with an unnoticed contamination event, or (ii) a double independent introduction of CCRAB (Cases 1 and 3) in UHRI. As already mentioned, Patient 3 was colonised simultaneously with OXA-181-producing CPEs, which are currently emerging in the Comoros archipelago, and could have acquired all the XDR bacteria abroad, including CCRAB. However, this patient had no recent history of travel outside Reunion Island. How Patient 5 was contaminated is not clear either. The positive sample collected in Nephrology medical ward suggests that he has been contaminated from the room environment, in which Patient 4 had stayed (Figure 1.a). The fact that all the patients had a negative rectal screening at their entry into the ICU supports the notion of a nosocomial outbreak. It also highlights the limitations of this screening procedure, which shows suboptimal sensitivity with A. baumannii, and which can be influenced by exposure to antibiotics [23-24]. Undoubtedly, the routes of diffusion of this XDR-AB clone into the ICU were complex and multiple. The early inter-unit transmission between Case 2 and Case 3 (in subunit A) points to the likely role of contaminated medical or personal equipment, as previously reported [25]. The occurrence of three strain resurgences in November 2019, March 2020 and June 2020 suggests that the strain persisted in the ICU environment. However, this could not be demonstrated despite the numerous samples collected in patients rooms, underscoring the fact that the detection of A. baumannii in the hospital environment may reveal challenging [26]. The sampling locations and frequency should indeed be optimised in order to maximise information on environmental contamination [27].

With regard to patient symptomatology, we noted a high rate of infections (61.5%; 8/13 cases) with a case-fatality rate of 46.2% (6 deaths out of 13 cases), greater than those usually reported for *A. baumannii* [11,28]. This might be explained by the lack of standardised treatments and recommendations to control these XDR microorganisms, which emerged recently [29–32].

This outbreak was caused by an MDR clone belonging to ICI, an epidemic lineage disseminated over the world [33]. Most of the MDR *A. baumannii* strains reported so far remained susceptible to polymyxins [34,35]. However, clonal expansion of a polymyxin resistant *A. baumannii* ST1 strains was recently observed in a Brazilian teaching hospital, highlighting the need for constant surveillance and implementation of adequate outbreak control measures [36]. Furthermore, it is worth noting that our clone is related to ST<sup>ox</sup>231, that was responsible for a large outbreak in hospitals of mainland France [37]. OXA-23/NDM-1 co-producing *A. baumannii* have been sporadically identified in some countries, but had never been reported from SIOA so far (Southern Africa, Mauritius, Madagascar). They would have emerged in the Comoros archipelago and Reunion Island (unpublished FNRC-AR data). Indeed, the first NDM-1/OXA-23 positive isolate (susceptible to colistin) was detected in 2015 in Mayotte island. Since then, 77% (n=95) of the 123 ST<sup>Pas</sup>1 OXA-23/NDM-1 producing strains collected by the FNRC-AR were isolated in this geographical area, or from patients who had travelled in this region of the world (FNRC-AR data). This finding is intriguing given the low consumption of antibiotics in Comoros archipelago, but might be due to increasing population exchanges with China and the Arabian Peninsula, where these two carbapenemases are prevalent [4,38,39].

Our study shows that Reunion Island has to face the import of XDR-bacteria from other islands and countries of the Indian Ocean zone. *De facto*, the UHRI acts as a sentinel for the survey of multidrug resistant bacteria disseminating in the SIOA [40]. This outbreak underscores the need for a systematic screening of all travellers coming to Reunion Island, and a special vigilance toward patients who have been medically evacuated.

As illustrated in this work, tracking of free-living nosocomial pathogens such as *A. baumannii* in the hospital environment may reveal difficult, even when thorough investigations are conducted. However, outbreaks due to such pathogens can be controlled by the early establishment of effective measures. In our hospital, XDR-AB have been reclassified as highly-resistant bacteria (such as CPE or Vancomycin-Resistant *Enterococcus*), and additional enhanced contact precautions are taken when new strains are isolated. Finally, because of the geographical remoteness of UHRI, high throughput sequencing methods need to be developed locally in order to rapidly investigate outbreaks due to XDR-bacteria.

#### 5. Conclusions

This study, which relates the emergence and intra-hospital spread of an XDR OXA-23/NDM-1 colistin-resistant *A. baumannii* clone, emphasises the need for strict application of prevention and infection control measures to control highly epidemic strains, which is a daily challenge in ICUs where patients require high and persistent care-loads. Because of their connection with mainland France, the two overseas territories of Reunion Island and Mayotte Island represent a potential source of contamination of Europe, by high-risk clones.

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#### **Declarations**

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- **Competing Interests**: None.
- Ethical Approval: This study was approved by the Ethics Committee of the French Society of
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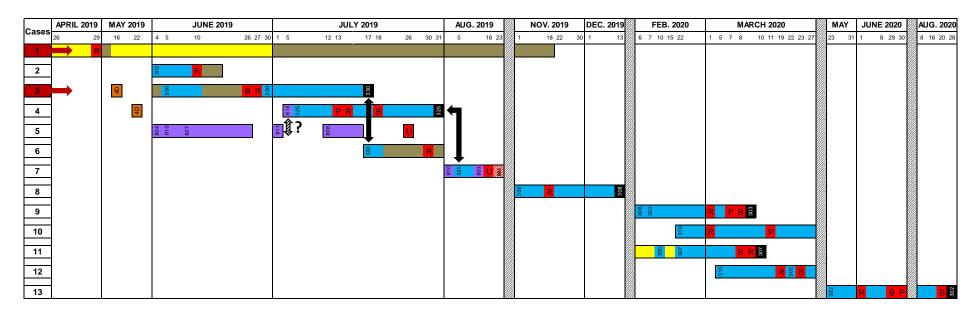
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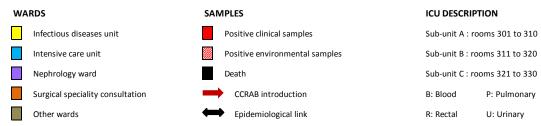
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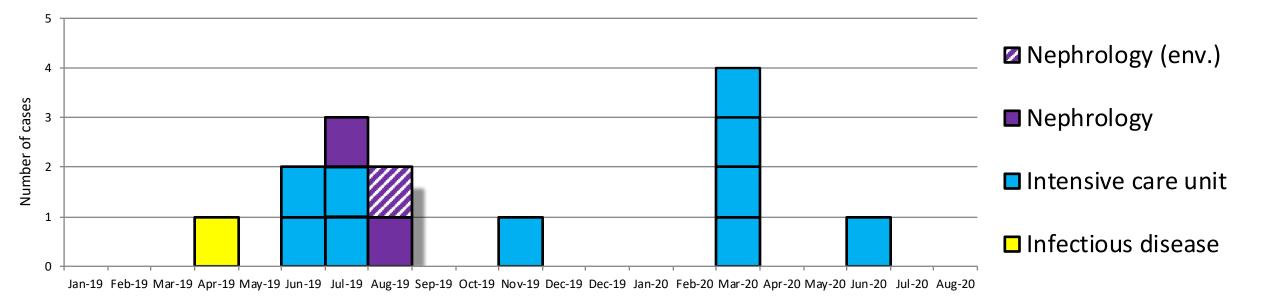
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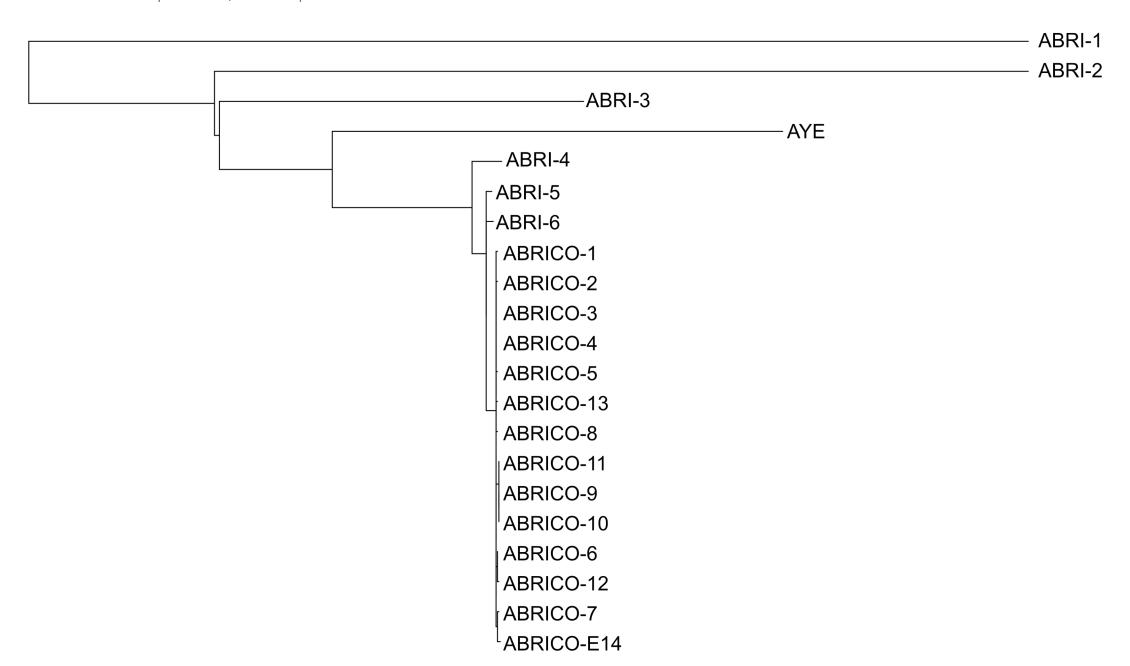
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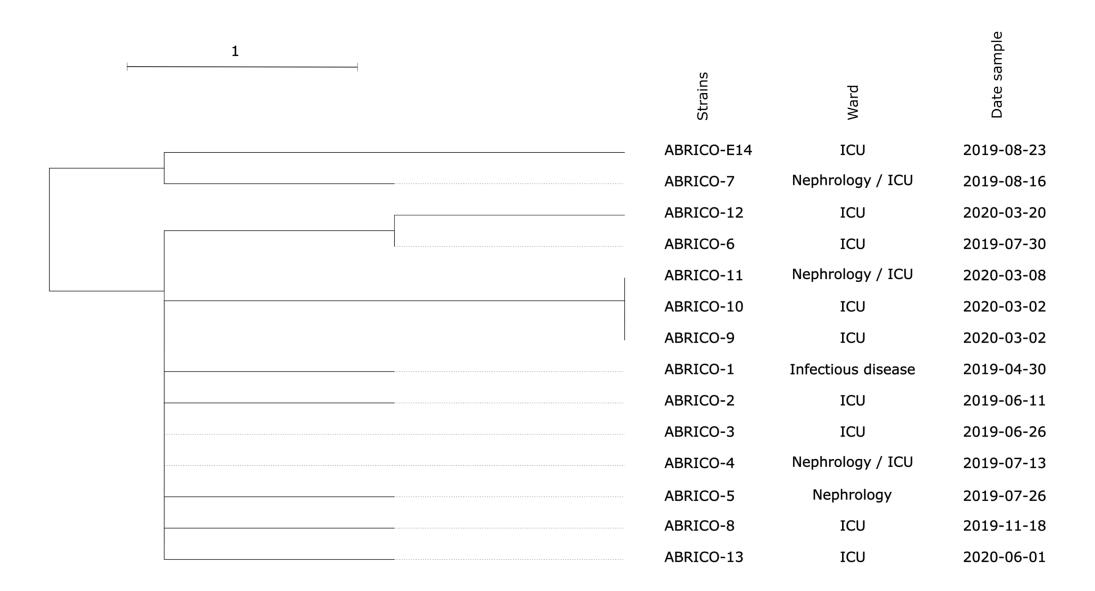
463 Figures 1.a and 1.b 464 1.a Timeline of patients colonised/infected with colistin- and carbapenem-resistant OXA-23/NDM-1-465 producing Acinetobacter baumannii hospitalised in the University Hospital of Reunion Island, Saint-466 Denis, France, April 2019-August 2020 (n=13). 467 Number in the box represents the number of the room in intensive care unit. 468 1.b Epidemic curve between January 2019 and August 2020. 469 Each box represents the first isolation of an XDR-Acinetobacter baumannii in a patient and the 470 corresponding hospitalisation ward. The purple hatched square represents the environmental isolate 471 detected in the Nephrology medical ward. 472 Figure 2.a and 2.b 473 474 2.a Phylogenetic analysis of colistin-resistant outbreak A. baumannii isolates and 7 other isolates 475 belonging to ST1 (Pasteur scheme) using the SNP from the "persistent" genes of PPanGGOLiN filtered 476 by Gubbins and tree constructed by RaXML-ng (model GTR+FO+IO; 100 bootstraps). The scale bar 477 represents the mean substitution rate by site. 478 2.b Phylogenetic analysis of the 14 CCRAB isolates using snippy-core, filtered and tree constructed 479 with Neighbor Joining algorithm. The scale bar represents the number of SNPs. 480











**Table 1.** Description of the XDR-*Acinetobacter baumannii* cases identified during the hospital outbreak investigation, Reunion Island, France, April 2019-August 2020 (n=13).

Outbreak case number	Isolate number	Age	Gender	Geographical origin	Hospitalisation ward	Sampling date	Specimen source	Infection/ Colonisation	Death	Epidemiological link
1	ABRICO-1	44	М	Mayotte	Infectious disease	Apr-2019	Rectal/Hip joint	Infection	No	No
2	ABRICO-2	74	F	Reunion Island	ICU	Jun-2019	Rectal	Colonisation	No	Yes
3	ABRICO-3	65	F	Reunion Island	ICU	Jun-2019	Blood	Infection	Yes	Yes
4	ABRICO-4	52	F	Reunion Island	Nephrology/ICU	Jul-2019	Rectal/Lung/Blood	Infection	Yes	Yes
5	ABRICO-5	64	M	Reunion Island	Nephrology	Jul-2019	Urine	Colonisation	No	No
6	ABRICO-6	68	M	Reunion Island	ICU	Jul-2019	Rectal	Colonisation	No	Yes
7	ABRICO-7	19	M	Reunion Island	Nephrology/ICU	Aug-2019	Urine	Colonisation	No	Yes
8	ABRICO-8	58	F	Reunion Island	ICU	Nov-2019	Rectal	Colonisation	Yes	Yes
9	ABRICO-9	48	M	Reunion Island	ICU	Mar-2020	Rectal/Blood	Infection	Yes	Yes
10	ABRICO-10	65	M	Reunion Island	ICU	Mar-2020	Rectal/Lung	Infection	No	Yes
11	ABRICO-11	47	M	Reunion Island	Infectious disease/ICU	Mar-2020	Rectal/Blood	Infection	Yes	Yes
12	ABRICO-12	73	M	Madagascar	ICU	Mar-2020	Rectal/Blood	Infection	No	Yes
13	ABRICO-13	43	М	Reunion Island	ICU	Jun-2020	Rectal/Lung/Blood	Infection	Yes	Yes

ICU: Intensive Care Unit

# **OUTBREAK DESCRIPTION**

University Hospital of Reunion Island



13 patients, including 11 in ICU



4 phases between Apr 2019 and Aug 2020



5 colonizations8 infections

→ 6 deaths (MR: 46.2%)



# **OUTBREAK CONTROL MEASURES**



Tri-sited simultaneous screening in ICU

Environmental sampling





Complete sequential rooms decontamination



Dedicated hygienist nurse (prevention of cross-transmission)

Multidisciplinary meeting (ICU's medical doctors, hygienists and microbiologists)

### STRAIN ANALYSIS

Colistin and carbapenem resistant Acinetobacter baumannii isolates

13 clinical + 1 environmental strains ST1<sup>Pas</sup>/ST<sup>Ox</sup>231

# 1-5 SNPs between the isolates

